Things fall apart: biological species form unconnected parsimony networks

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The generality of operational species definitions is limited by problematic definitions of between-species divergence. A recent phylogenetic species concept based on a simple objective measure of statistically significant genetic differentiation uses between-species application of statistical parsimony networks that are typically used for population genetic analysis within species. Here we review recent phylogeographic studies and reanalyse several mtDNA barcoding studies using this method. We found that (i) alignments of DNA sequences typically fall apart into a separate subnetwork for each Linnean species (but with a higher rate of true positives for mtDNA data) and (ii) DNA sequences from single species typically stick together in a single haplotype network. Departures from these patterns are usually consistent with hybridization or cryptic species diversity.

Keywords: phylogenetic species; phylogeography; Astraptes; Cypraeidae

1. INTRODUCTION
The interruption of gene flow by speciation events is widely expected to produce large observed genetic discontinuities that can be used as operational definitions of species or evolutionarily significant units (ESUs; Mallet 1995; Sites & Marshall 2003, 2004; Vogler & Monaghan 2007). For example, the Barcodes of Life Initiative depends on the measurement of statistically significant genetic differentiation uses between-species application of statistical parsimony networks that are typically used for population genetic analysis within species. Here we review recent phylogeographic studies and reanalyse several mtDNA barcoding studies using this method. We found that (i) alignments of DNA sequences typically fall apart into a separate subnetwork for each Linnean species (but with a higher rate of true positives for mtDNA data) and (ii) DNA sequences from single species typically stick together in a single haplotype network. Departures from these patterns are usually consistent with hybridization or cryptic species diversity.

2. MATERIAL AND METHODS
We identified from the Science Citation Index Expanded (up to 4 October 2006) a total of 727 publications that cited Clement et al. (2000) and used the statistical parsimony method as implemented in the software application TCS (see electronic supplementary material). We reviewed the results of 517 of these studies (table S1) that were available through our home institution library, included an unambiguous number of parsimony networks and taxa, and used the 95% probability of parsimony. For each network analysis \((n=663)\), we noted the number of taxa and 95% subnetworks. In cases where the parsimony network results led the authors to reinterpret species and ESU designations, we used this revised number of taxa. We also reanalysed data from recent studies of butterflies (Hebert et al. 2004) and marine snails (Cypraeidae or cowries; Meyer & Paulay 2005) related to the mtDNA barcoding approach to documenting biodiversity. We produced 95% parsimony networks from these mtDNA sequence alignments in TCS v. 1.21. We noted the number of subnetworks and taxa, and the 95% connection limit \((L_{95})\). We used receiver-operating characteristic (ROC) analysis (Metz 2006) to assess the diagnostic accuracy of TCS networks to differentiate species (see definitions in the electronic supplementary material).

3. RESULTS

(a) Review
Most of the 663 published network analyses (78%) found the same number of subnetworks as taxa and correctly identified all taxa (figure 1a). The mean \((\pm\text{s.e.})\) rate of false positives among all analyses was \(0.006 \pm 0.002\) (few errors were made in joining all the members of a sampled species into one subnetwork).

The rate of true positives was only measurable for studies in which more than one species was used in a network analysis. Among 192 such analyses (34%), the mean rate of true positives was 0.521 \pm 0.034 (only about half of expected species boundaries were identified).

In 36 analyses, authors attributed apparent discordance between networks and taxa (6 false positives and 80 false negatives) to hybridization (e.g. Kronforst et al. 2006). Other sources of false positives could be inadequate sampling (e.g. figure S1), historical vicariance with rapid lineage sorting or cryptic speciation with undocumented phenotypic divergence (Knowlton 1993).

Failures to detect true positives (i.e. incidents of false negatives) may be caused by limited genetic divergence and incomplete lineage sorting between sister taxa (e.g. figure S2), or by selective constraints on lineage divergence, such as studies that used conserved, expressed nuclear genes.

Among 24 analyses of mtDNA COI sequences (the standard genetic marker in animal barcoding studies; Hebert et al. 2003; figure 1b) in two or more species, the mean false-positive rate was 0.008 \pm 0.006. The mean true-positive rate was considerably higher (0.916 \pm 0.046) than the rate for all studies (0.521).

(b) Barcoding: butterflies
An early and influential barcoding study (Hebert et al. 2004) suggested at least 10 cryptic species in the Astraptes fulgerator complex to which the authors gave provisional acronym identifiers based on larval morphology and food plant use. Statistical parsimony analysis of the mtDNA sequence alignment produced eight subnetworks. Seven of these subnetworks and quantitatively reliable across a broad range of taxa and genetic markers.
Altogether, the statistical parsimony method correctly identified all of the well-corroborated cryptic species in the A. fulgerator complex, and failed to differentiate the FABOV, INGCUP and HIHAMP phenotypes, probably because they are either conspecific or very recently speciated.

**4. DISCUSSION**

Two recent steps towards a statistically objective phylogenetic species concept have used bifurcating trees in a maximum likelihood or Bayesian framework. Matz & Nielsen (2005) and Nielsen & Matz (2006) developed a likelihood ratio test for inclusion of a test sequence in a single species sample. Pons et al. (2006) identified the boundary between two species as a shift from short branches within taxa (caused by coalescence processes in populations) to long branches between taxa (caused by speciation and extinction events), analogous to the difference between within- and among-species lineage sorting evident in statistical parsimony networks.

Recent analyses of specific taxa (cited above) show that the 95% parsimony connection limit can provide an additional and simple quantitative standard for phylogenetic species (Monaghan et al. 2006). Our review and reanalyses suggest that this standard has a low rate of false-positive errors across a broad range of taxa, speciation problems and genetic markers, and that it may be generally useful for assigning unknown specimens to known, well-sampled taxa (DNA barcoding as defined by Vogler & Monaghan 2007). This approach might be particularly useful for barcoding studies in which morphological or ecological species markers are labile (such as caterpillar host plant use by INGCUP and FABOV forms of A. fulgerator; Hebert et al. 2004). The parsimony connection limit appears to have a higher true-positive rate for successful identification of known species boundaries, and by extension for discovering new cryptic species from sequence data (DNA taxonomy; Vogler & Monaghan 2007), when applied to non-recombining loci with
Table 1. Sensitivity analyses of parsimony network results for two cowrie genera in which errors can (Palmadusta) or cannot (Cribrarula) be reduced to zero by choosing a probability of parsimony different from the default value of 0.95 (bold); and summed across 15 well-sampled genera (Bistolida, Cribrarula, Cypracovula, Erosaria, Erronea, Leporicyprea, Luria, Lyncina, Mauritia, Monetaria, Notadusta, Ovatipsa, Palmadusta, Pustularia and Zoila) that included one or more false-negative ESUs or false-positive subnetworks. (No errors were found in four additional genera (Cypraea, Notocypraea, Talostolida and Purpuradusta). Adapted from Meyer & Paulay (2005).)

<table>
<thead>
<tr>
<th>group</th>
<th>probability of parsimony</th>
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<tbody>
<tr>
<td></td>
<td>0.90</td>
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<tr>
<td><strong>Palmadusta</strong> (16 ESUs)</td>
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<tr>
<td>false negatives</td>
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<tr>
<td>false positives</td>
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<tr>
<td>total errors</td>
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</tr>
<tr>
<td><strong>Cribrarula</strong> (18 ESUs)</td>
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<tr>
<td><strong>15 genera (178 ESUs)</strong></td>
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</tr>
<tr>
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<td>57</td>
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Morando, M., Avila, L. J. & Sites, J. W. 2003 Sampling strategies for delimiting species: genes, individuals, and populations in the *Liothaemus elongate–kriegi* complex...


